



INVESTOR IN PEOPLE

CERTIFIED COPY OF PRIORITY DOCUMENT

The Patent Office
Concept House
Cardiff Road
Newport
South Wales
NP10 8QQ



I, the undersigned, being an officer duly authorised in accordance with Section 74(1) and (4) of the Deregulation & Contracting Out Act 1994, to sign and issue certificates on behalf of the Comptroller-General, hereby certify that annexed hereto is a true copy of the documents as originally filed in connection with the patent application identified therein.

I also certify that by virtue of an assignment registered under the Patents Act 1977, the application is now proceeding in the name as substituted.

In accordance with the Patents (Companies Re-registration) Rules 1982, if a company named in this certificate and any accompanying documents has re-registered under the Companies Act 1980 with the same name as that with which it was registered immediately before re-registration save for the substitution as, or inclusion as, the last part of the name of the words "public limited company" or their equivalents in Welsh, references to the name of the company in this certificate and any accompanying documents shall be treated as references to the name with which it is so re-registered.

In accordance with the rules, the words "public limited company" may be replaced by p.l.c., plc, P.L.C. or PLC.

Re-registration under the Companies Act does not constitute a new legal entity but merely subjects the company to certain additional company law rules.

Signed *Andrews*.

Dated 7 December 2000

BEST AVAILABLE COPY

GB0000992.8

By virtue of a direction given under Section of the Patents Act 1977, the application is proceeding in the name of

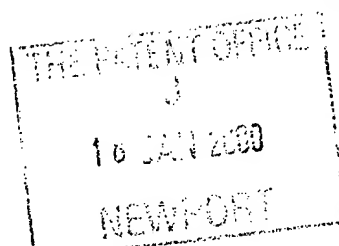
ASTRAZENECA AB,
Incorporated in Sweden,
S-151 85 Sodertalje,
Sweden

[ADP No. 07822448003]

1/77

Request for grant of a patent

(See the notes on the back of this form. You can also get an explanatory leaflet from the Patent Office to help you fill in this form)



The Patent Office

Cardiff Road
Newport
Gwent NP9 1RH

1. Your reference

PHM 99-191

2. Patent application number

(The Patent Office will fill in this part)

0000992.8

18 JAN 2000

3. Full name, address and postcode of the or of each applicant (underline all surnames)

AstraZeneca UK Limited
15 Stanhope Gate
LONDON
W1Y 6LN, GB

Patents ADP number (if you know it)

6254007002

If the applicant is a corporate body, give the country/state of its incorporation

7810294001

4. Title of the invention

METHODS

5. Name of your agent (if you have one)

GILES, Allen, Frank

"Address for service" in the United Kingdom to which all correspondence should be sent (including the postcode)

ZENECA Pharmaceuticals
Intellectual Property Department
Mereside, Alderley Park,
Macclesfield, Cheshire, Sk10 4TG, GB

44700

Patents ADP number (if you know it)

6988463001

6. If you are declaring priority from one or more earlier patent applications, give the country and the date of filing of the or of each of these earlier applications and (if you know it) the or each application number

Country

Priority application number
(if you know it)

Date of filing
(day / month / year)

7. If this application is divided or otherwise derived from an earlier UK application, give the number and the filing date of the earlier application

Number of earlier application

Date of filing
(day / month / year)

8. Is a statement of inventorship and of right to grant of a patent required in support of this request? (Answer 'Yes' if:

- a) any applicant named in part 3 is not an inventor, or
 - b) there is an inventor who is not named as an applicant, or
 - c) any named applicant is a corporate body.
- See note (d))

9. Enter the number of sheets for any of the following items you are filing with this form. Do not count copies of the same document

Continuation sheets of this form

Description

19

Claim(s)

Abstract

Drawing(s)

10. If you are also filing any of the following, state how many against each item.

Priority documents

Translations of priority documents

Statement of inventorship and right to grant of a patent (Patents Form 7/77)

Request for preliminary examination and search (Patents Form 9/77)

Request for substantive examination (Patents Form 10/77)

Any other documents (please specify)

11.

I/We request the grant of a patent on the basis of this application.

Signature

Lynda M. Slack Date 17th Jan 2000.

12. Name and daytime telephone number of person to contact in the United Kingdom

Mrs Lynda May Slack - 01625 516173

Warning

After an application for a patent has been filed, the Comptroller of the Patent Office will consider whether publication or communication of the invention should be prohibited or restricted under Section 22 of the Patents Act 1977. You will be informed if it is necessary to prohibit or restrict your invention in this way. Furthermore, if you live in the United Kingdom, Section 23 of the Patents Act 1977 stops you from applying for a patent abroad without first getting written permission from the Patent Office unless an application has been filed at least 6 weeks beforehand in the United Kingdom for a patent for the same invention and either no direction prohibiting publication or communication has been given, or any such direction has been revoked.

Notes

- If you need help to fill in this form or you have any questions, please contact the Patent Office on 0645 500505.
- Write your answers in capital letters using black ink or you may type them.
- If there is not enough space for all the relevant details on any part of this form, please continue on a separate sheet of paper and write "see continuation sheet" in the relevant part(s). Any continuation sheet should be attached to this form.
- If you have answered 'Yes' Patents Form 7/77 will need to be filed.
- Once you have filled in the form you must remember to sign and date it.
- For details of the fee and ways to pay please contact the Patent Office.

METHODS

This invention relates to polymorphisms in the human pyruvate dehydrogenase complex E2 (PDH E2 or PDC E2) gene and corresponding novel allelic polypeptides encoded thereby. The invention also relates to methods and materials for analysing allelic variation in the PDH E2 gene, and to the use of PDH E2 polymorphism in the diagnosis and treatment of diseases in which modulation of pyruvate dehydrogenase activity could be of therapeutic benefit, such as diabetes, asthma, obesity, sepsis and peripheral vascular disease.

The production of energy for biosynthesis of complex molecules and for muscle contraction is mediated by the hydrolysis of high energy phosphate bonds within adenosine triphosphate (ATP). In oxidative metabolism ATP is generated from acetyl coenzyme A (acetyl CoA), which itself is produced by the beta-oxidation of fatty acids, or as a result of the metabolism of glucose via the glycolytic pathway. The key regulatory enzyme which controls the rate of acetyl CoA formation from glucose is pyruvate dehydrogenase (PDH), which catalyses the oxidation of pyruvate to acetyl CoA and carbon dioxide with concomitant reduction of NAD to NADH.

PDH is a multienzyme complex located in the mitochondrial matrix, containing multiple copies of three enzyme components required to complete the conversion of pyruvate to acetyl CoA (Patel and Roche 1990; FASEB J., 4: 3224-3233). E1 (pyruvate decarboxylase, E.C. 1.2.4.1) catalyses the non-reversible removal of carbon dioxide from pyruvate; E2 (dihydrolipoamide acetyltransferase, E.C. 2.3.1.12) catalyses the formation of acetyl CoA; and E3 (dihydrolipoamide dehydrogenase, E.C. 1.8.1.4) reduces NAD to NADH. The E1 enzyme is a heterotetramer composed of two α and two β subunits. Decarboxylation of pyruvate, catalysed by E1 is the rate limiting step in the overall activity of the PDH complex. This step is also the target for a cycle of phosphorylation and dephosphorylation which forms one of the main mechanisms for regulating PDH activity. Two additional enzyme activities are also associated with the PDH complex: a specific kinase (PDK) which is capable of phosphorylating E1 α at three serine residues, and a loosely-associated specific phosphatase which reverses the phosphorylation. Phosphorylation of only one of the three serine residues on E1 α renders E1 inactive. Removal of the phosphate groups by the specific phosphatase

restores activity. Thus, the proportion of PDH in its active (dephosphorylated) state is determined by a balance between the activity of the kinase and phosphatase. The activity of the kinase may be regulated in vivo by the relative concentrations of metabolic substrates such as NAD/NADH, CoA/acetylCoA and ADP/ATP as well as by the availability of pyruvate
5 itself, therefore providing highly regulated, responsive control of PDH activity.

Genetic abnormalities in the PDH complex are the most common cause of primary lactic acidosis in humans. The majority of cases have been linked with a defect in the E1 α subunit. Pathologies associated with defects in the PDH complex conform to a broad clinical spectrum ranging from fatal lactic acidosis in the newborn, to a range of chronic
10 neurodegenerative conditions with gross structural abnormalities in the central nervous system. E1 α deficiency is an X-linked disorder which manifests different patterns of clinical presentation between males and females. In addition, heterozygous females show a wide variation in clinical severity of the disease, due largely to variations in the pattern of X-inactivation and differential effects of specific gene mutations on the expression, stability and
15 activity of the mutant protein. A number of mutations in the PDH gene which lead to pyruvate dehydrogenase deficiency have been documented (for a review see NIH OMIM database, reference 312170).

In disease states such as both non-insulin dependent (NIDDM) and insulin-dependent diabetes (IDDM), oxidation of lipids is increased with a concomitant reduction in utilisation
20 of glucose, contributing to the hyperglycaemia. The activity of PDH is reduced in both insulin-dependent and non insulin-dependent diabetes. A further consequence of reduced PDH activity is an increase in pyruvate concentration resulting in increased availability of lactate as a substrate for hepatic gluconeogenesis. Diabetes is further exacerbated by impaired insulin secretion, which has been shown to be associated with reduced PDH activity in pancreatic β -
25 cells. It is believed that increasing the activity of PDH may increase the rate of glucose oxidation and hence overall glucose utilisation, in addition to reducing hepatic glucose output.

Oxidation of glucose is capable of yielding more molecules of ATP per mole of oxygen than is oxidation of fatty acids, therefore in conditions where energy demand may exceed energy supply, such as myocardial ischaemia and reperfusion, intermittent
30 claudication, cerebral ischaemia and reperfusion, shifting the balance of substrate utilisation in

favour of glucose metabolism may be expected to improve the ability to maintain ATP levels and hence function. Activation of PDH is predicted to have this effect.

An agent which is capable of activating PDH is expected to be of benefit in treating conditions where an excess of circulating lactic acid is manifest such as in certain cases of
5 sepsis.

The agent dichloroacetic acid which increases the activity of PDH after acute administration in animals (Vary et al., 1988; Circ. Shock, 24: 3-18) has been shown to have the predicted effects in reducing glycaemia (Stacpoole et al, 1978 N. Engl. J. Med. 298, 526-530) and as a therapy for myocardial ischaemia (Bersin and Stacpoole 1997; American Heart
10 Journal, 134: 841-855) and lactic acidemia (Stacpoole et al, 1983 N. Engl. J. Med 309, 390-396).

cDNA sequences encoding PDH E2 have been submitted to public databases under the following accession numbers: Y00978, J03866. We believe that sequence J03866 contains a number of errors, discussed in more detail in Example 2. All positions in the human PDH E2
15 gene herein refer to the positions in EMBL accession number Y00978 unless stated otherwise or apparent from the context.

DNA polymorphisms may lead to variations in amino acid sequence and consequently to altered protein structure and functional activity. Polymorphisms may also affect mRNA synthesis, maturation, transportation and stability. Polymorphisms which do not result in
20 amino acid changes (silent polymorphisms) or which do not alter any known consensus sequences may nevertheless have a biological effect, for example by altering mRNA folding or stability.

Knowledge of polymorphisms may be used to help identify patients most suited to therapy with particular pharmaceutical agents (this is often termed "pharmacogenetics") .
25 Pharmacogenetics can also be used in pharmaceutical research to assist the drug selection process. Polymorphisms may be used in mapping the human genome and to elucidate the genetic component of diseases. The reader is directed to the following references for background details on pharmacogenetics and other uses of polymorphism detection: Linder *et al.* (1997), Clinical Chemistry, 43, 254; Marshall (1997), Nature Biotechnology, 15, 1249;

International Patent Application WO 97/40462, Spectra Biomedical; and Schafer *et al.* (1998), Nature Biotechnology, 16, 33.

Clinical trials have shown that patient response to treatment with pharmaceuticals is often heterogeneous. Thus there is a need for improved approaches to pharmaceutical agent
5 design and therapy.

Variations in polypeptide sequence will be referred to as follows: original amino acid (using one or three letter nomenclature), position, new amino acid. For (a hypothetical) example "D25K" or "Asp25Lys" means that at position 25 an aspartic acid (D) has been changed to lysine (K). Multiple variations in one polypeptide will be shown between square
10 brackets with individual variations separated by commas.

The present invention is based on the discovery of two single nucleotide polymorphisms (SNPs) in the coding region of the human PDH E2 gene.

According to one aspect of the present invention there is provided a method for the diagnosis of a polymorphism in a PDH E2 gene in a human, which method comprises
15 determining the sequence of the nucleic acid of the human at one or more of positions 857 and 1255 in the PDH E2 gene as defined by the positions in EMBL accession number Y00978; and determining the status of the human by reference to polymorphism in the PDH E2 gene.

The term human includes both a human having or suspected of having a PDH-mediated disease and an asymptomatic human who may be tested for predisposition or
20 susceptibility to such disease. At each position the human may be homozygous for an allele or the human may be a heterozygote.

The term 'PDH-mediated disease' means any disease in which changing the level of PDH or changing the activity of PDH would be of therapeutic benefit.

The term 'PDH drug' means any drug which changes the level of PDH or changes the
25 activity of PDH. A drug which increases the activity of PDH is preferred.

The term polymorphism includes single nucleotide substitution, nucleotide insertion and nucleotide deletion, which in the case of insertion and deletion includes insertion or deletion of one or more nucleotides at a position of a gene and variable numbers of a repeated DNA sequence.

In one embodiment of the invention preferably the method for diagnosis described herein is one in which the single nucleotide polymorphism at position 857 in the PDH E2 gene as defined by the positions in EMBL accession number Y00978 is presence of T and/or C.

- 5 In another embodiment of the invention preferably the method for diagnosis described herein is one in which the single nucleotide polymorphism at position 1255 in the PDH E2 gene as defined by the positions in EMBL accession number Y00978 is presence of G and/or A.

The method for diagnosis is preferably one in which the sequence is determined by a
10 method selected from amplification refractory mutation system and restriction fragment length polymorphism.

In another aspect of the invention we provide a method for the diagnosis of PDH-mediated disease, which method comprises:

- i) obtaining sample nucleic acid from an individual,
- 15 ii) detecting the presence or absence of a variant nucleotide at one or more of positions 857 and 1255 in the PDH E2 gene as defined by the positions in EMBL accession number Y00978; and
- iii) determining the status of the individual by reference to polymorphism in the PDH E2 gene.

20 Allelic variation at position 857 in the PDH E2 gene as defined by the positions in EMBL accession number Y00978 consists of a single base substitution from T (the published base), preferably to C. Allelic variation at position 1255 in the PDH E2 gene as defined by the positions in EMBL accession number Y00978 consists of a single base substitution from G (the published base), preferably to A. The status of the individual may be determined by
25 reference to allelic variation at any one or both positions optionally in combination with any other polymorphism in the gene that is (or becomes) known.

The test sample of nucleic acid is conveniently present in a sample of blood, sputum, skin, bronchoalveolar lavage fluid, or other body fluid or tissue obtained from an individual. It will be appreciated that the test sample may equally comprise a nucleic acid sequence
30 corresponding to the sequence in the test sample, that is to say that all or a part of the region

in the sample nucleic acid may firstly be amplified using any convenient technique e.g. PCR, before analysis of allelic variation.

It will be apparent to the person skilled in the art that there are a large number of analytical procedures which may be used to detect the presence or absence of variant nucleotides at one or more polymorphic positions of the invention. In general, the detection of allelic variation requires a mutation discrimination technique, optionally an amplification reaction and optionally a signal generation system. Table 1 lists a number of mutation detection techniques, some based on PCR. These may be used in combination with a number of signal generation systems, a selection of which is listed in Table 2. Further amplification techniques are listed in Table 3. Many current methods for the detection of allelic variation are reviewed by Nollau *et al.*, Clin. Chem. 43, 1114-1120, 1997; and in standard textbooks, for example "Laboratory Protocols for Mutation Detection", Ed. by U. Landegren, Oxford University Press, 1996 and "PCR", 2nd Edition by Newton & Graham, BIOS Scientific Publishers Limited, 1997.

15

Abbreviations:

ALEX TM	Amplification refractory mutation system linear extension
APEX	Arrayed primer extension
ARMST TM	Amplification refractory mutation system
b-DNA	Branched DNA
CMC	Chemical mismatch cleavage
bp	base pair
COPS	Competitive oligonucleotide priming system
DGGE	Denaturing gradient gel electrophoresis
FRET	Fluorescence resonance energy transfer
IDDM	Insulin-dependent diabetes mellitus
LCR	Ligase chain reaction
MASDA	Multiple allele specific diagnostic assay
NASBA	Nucleic acid sequence based amplification

NIDDM	non-insulin dependent diabetes mellitus
OLA	Oligonucleotide ligation assay
PCR	Polymerase chain reaction
PDH	Pyruvate Dehydrogenase
PDK	Pyruvate Dehydrogenase Kinase
PDK2	Pyruvate Dehydrogenase Kinase Isoenzyme 2
PTT	Protein truncation test
RFLP	Restriction fragment length polymorphism
SDA	Strand displacement amplification
SNP	Single nucleotide polymorphism
SSCP	Single-strand conformation polymorphism analysis
SSR	Self sustained replication
TGGE	Temperature gradient gel electrophoresis
3'UTR	3' Untranslated Region

Table 1 - Mutation Detection Techniques

General: DNA sequencing, Sequencing by hybridisation

Scanning: PTT*, SSCP, DGGE, TGGE, Cleavase, Heteroduplex analysis, CMC, Enzymatic

5 mismatch cleavage

* Note: not useful for detection of promoter polymorphisms.

Hybridisation Based:

Solid phase hybridisation: Dot blots, MASDA, Reverse dot blots, Oligonucleotide arrays (DNA Chips).

10 Solution phase hybridisation: Taqman™ - US-5210015 & US-5487972 (Hoffmann-La Roche), Molecular Beacons - Tyagi *et al* (1996), Nature Biotechnology, **14**, 303; WO 95/13399 (Public Health Inst., New York).

Extension Based: ARMST™, ALEX™ - European Patent No. EP 332435 B1 (Zeneca Limited), COPS - Gibbs *et al* (1989), Nucleic Acids Research, **17**, 2347.

15 **Incorporation Based:** Mini-sequencing, APEX

Restriction Enzyme Based: RFLP, Restriction site generating PCR

Ligation Based: OLA

Other: Invader assay

5 Table 2 - Signal Generation or Detection Systems

Fluorescence: FRET, Fluorescence quenching, Fluorescence polarisation - United Kingdom Patent No. 2228998 (Zeneca Limited)

Other: Chemiluminescence, Electrochemiluminescence, Raman, Radioactivity, Colorimetric, Hybridisation protection assay, Mass spectrometry.

10

Table 3 - Further Amplification Methods

SSR, NASBA, LCR, SDA, b-DNA

Preferred mutation detection techniques include ARMSTTM, ALEXTM, COPS, Taqman,
15 Molecular Beacons, RFLP, and restriction site based PCR and FRET techniques.

Particularly preferred methods include ARMSTTM and RFLP based methods. ARMSTTM is an especially preferred method.

In a further aspect, the diagnostic methods of the invention are used to assess the efficacy of therapeutic compounds in the treatment of PDH-mediated diseases such as
20 diabetes, asthma, obesity, sepsis, and peripheral vascular disease.

Assays, for example reporter-based assays, may be devised to detect whether one or more of the above polymorphisms affect transcription levels and/or message stability.

Individuals who carry particular allelic variants of the PDH E2 gene may therefore exhibit differences in their ability to regulate protein biosynthesis under different
25 physiological conditions and may display altered abilities to react to different diseases. In addition, differences in protein regulation arising as a result of allelic variation may have a direct effect on the response of an individual to drug therapy. The diagnostic methods of the invention may be useful both to predict the clinical response to such agents and to determine therapeutic dose.

In a further aspect, the diagnostic methods of the invention, are used to assess the predisposition of an individual to diseases mediated by PDH. This may be particularly relevant in the development of diabetes, asthma, obesity, sepsis, and peripheral vascular disease and other diseases which are mediated by PDH. The present invention may be used to
5 recognise individuals who are particularly at risk from developing these conditions.

Low frequency polymorphisms may be particularly useful for haplotyping as described below. A haplotype is a set of alleles found at linked polymorphic sites (such as within a gene) on a single (paternal or maternal) chromosome. If recombination within the gene is random, there may be as many as 2^n haplotypes, where 2 is the number of alleles at
10 each polymorphic position and n is the number of polymorphic positions. One approach to identifying mutations or polymorphisms which are correlated with clinical response is to carry out an association study using all the haplotypes that can be identified in the population of interest. The frequency of each haplotype is limited by the frequency of its rarest allele, so that polymorphisms with low frequency alleles are particularly useful as markers of low
15 frequency haplotypes. As particular mutations or polymorphisms associated with certain clinical features, such as adverse or abnormal events, are likely to be of low frequency within the population, low frequency polymorphisms may be particularly useful in identifying these mutations (for examples see: De Stefano V et al., *Ann Hum Genet* (1998) 62:481-90; and Keightley AM et al., *Blood* (1999) 93:4277-83.

20 In a further aspect, the diagnostic methods of the invention are used in the development of new drug therapies which selectively target one or more allelic variants of the PDH E2 gene. Identification of a link between a particular allelic variant and predisposition to disease development or response to drug therapy may have a significant impact on the design of new drugs. Drugs may be designed to regulate the biological activity of variants
25 implicated in the disease process whilst minimising effects on other variants.

In a further diagnostic aspect of the invention the presence or absence of variant nucleotides is detected by reference to the loss or gain of, optionally engineered, sites recognised by restriction enzymes.

According to another aspect of the present invention there is provided a nucleic acid
30 comprising any one of the following polymorphisms:

the nucleic acid of EMBL accession number Y00978 with C at position 857 as defined by the positions in EMBL accession number Y00978;

the nucleic acid of EMBL accession number Y00978 with A at position 1255 as defined by the position in EMBL accession number Y00978;

- 5 or a complementary strand thereof or an antisense sequence thereto or a fragment thereof of at least 20 bases comprising at least one polymorphism.

Fragments are at least 17 bases, more preferably at least 20 bases, more preferably at least 30 bases.

The scope of the invention does not extend to any nucleic acid as it is found in nature.

- 10 A nucleic acid of the invention is preferably in isolated form, for example through being at least partially purified from any substance with which it occurs naturally (if any).

Novel sequence disclosed herein, may be used in another embodiment of the invention to regulate expression of the gene in cells by the use of antisense constructs. To enable methods of down-regulating expression of the gene of the present invention in

15 mammalian cells, an example antisense expression construct can be readily constructed for instance using the pREP10 vector (Invitrogen Corporation). Transcripts are expected to inhibit translation of the gene in cells transfected with this type of construct. Antisense transcripts are effective for inhibiting translation of the native gene transcript, and capable of inducing the effects (e.g., regulation of tissue physiology) herein described. Oligonucleotides

20 which are complementary to and hybridisable with any portion of novel gene mRNA disclosed herein are contemplated for therapeutic use. U.S. Patent No. 5,639,595, "Identification of Novel Drugs and Reagents", issued Jun. 17, 1997, wherein methods of identifying oligonucleotide sequences that display in vivo activity are thoroughly described, is herein incorporated by reference. Expression vectors containing random oligonucleotide

25 sequences derived from previously known polynucleotides are transformed into cells. The cells are then assayed for a phenotype resulting from the desired activity of the oligonucleotide. Once cells with the desired phenotype have been identified, the sequence of the oligonucleotide having the desired activity can be identified. Identification may be accomplished by recovering the vector or by polymerase chain reaction (PCR) amplification

30 and sequencing the region containing the inserted nucleic acid material. Antisense molecules

can be synthesised for antisense therapy. These antisense molecules may be DNA, stable derivatives of DNA such as phosphorothioates or methylphosphonates, RNA, stable derivatives of RNA such as 2'-O-alkylRNA, or other oligonucleotide mimetics. U.S. Patent No. 5,652,355, "Hybrid Oligonucleotide Phosphorothioates", issued July 29, 1997, and U.S.

5 Patent No. 5,652,356, "Inverted Chimeric and Hybrid Oligonucleotides", issued July 29, 1997, which describe the synthesis and effect of physiologically-stable antisense molecules, are incorporated by reference. Antisense molecules may be introduced into cells by microinjection, liposome encapsulation or by expression from vectors harboring the antisense sequence.

10 The invention further provides nucleotide primers which can detect the polymorphisms of the invention.

According to another aspect of the present invention there is provided an allele specific primer capable of detecting a PDH E2 gene polymorphism at one or more of positions 857 and 1255 in the PDH E2 gene as defined by the positions in EMBL accession number
15 Y00978.

An allele specific primer is used, generally together with a constant primer, in an amplification reaction such as a PCR reaction, which provides the discrimination between alleles through selective amplification of one allele at a particular sequence position e.g. as used for ARMS™ assays. The allele specific primer is preferably 17- 50 nucleotides, more
20 preferably about 17-35 nucleotides, more preferably about 17-30 nucleotides.

An allele specific primer preferably corresponds exactly with the allele to be detected but derivatives thereof are also contemplated wherein about 6-8 of the nucleotides at the 3' terminus correspond with the allele to be detected and wherein up to 10, such as up to 8, 6, 4, 2, or 1 of the remaining nucleotides may be varied without significantly affecting the
25 properties of the primer.

Primers may be manufactured using any convenient method of synthesis. Examples of such methods may be found in standard textbooks, for example "Protocols for Oligonucleotides and Analogues; Synthesis and Properties," Methods in Molecular Biology Series; Volume 20; Ed. Sudhir Agrawal, Humana ISBN: 0-89603-247-7; 1993; 1st Edition. If
30 required the primer(s) may be labelled to facilitate detection.

According to another aspect of the present invention there is provided an allele-specific oligonucleotide probe capable of detecting a PDH E2 gene polymorphism at one or more of positions 857 and 1255 in the PDH E2 gene as defined by the positions in EMBL accession number Y00978.

- 5 The allele-specific oligonucleotide probe is preferably 17- 50 nucleotides, more preferably about 17-35 nucleotides, more preferably about 17-30 nucleotides.

The design of such probes will be apparent to the molecular biologist of ordinary skill. Such probes are of any convenient length such as up to 50 bases, up to 40 bases, more conveniently up to 30 bases in length, such as for example 8-25 or 8-15 bases in length. In
10 general such probes will comprise base sequences entirely complementary to the corresponding wild type or variant locus in the gene. However, if required one or more mismatches may be introduced, provided that the discriminatory power of the oligonucleotide probe is not unduly affected. The probes of the invention may carry one or more labels to facilitate detection.

- 15 According to another aspect of the present invention there is provided a diagnostic kit comprising an allele specific oligonucleotide probe of the invention and/or an allele-specific primer of the invention.

The diagnostic kits may comprise appropriate packaging and instructions for use in the methods of the invention. Such kits may further comprise appropriate buffer(s), nucleotides,
20 and polymerase(s) such as thermostable polymerases, for example taq polymerase.

In another aspect of the invention, the polymorphisms of this invention may be used as genetic markers in linkage studies. This particularly applies to the polymorphism at position 1255 in the PDH E2 gene as defined by the positions in EMBL accession number Y00978 because of its relatively high frequency (see Examples below).

- 25 According to another aspect of the present invention there is provided a method of treating a human in need of treatment with a PDH drug in which the method comprises:
- i) diagnosis of a polymorphism in the PDH E2 gene in the human, which diagnosis comprises determining the sequence of the nucleic acid at one or more of positions 857 and 1255 in the PDH E2 gene as defined by the positions in EMBL accession number Y00978,

and determining the status of the human by reference to polymorphism in the PDH E2 gene;
and

ii) administering an effective amount of a PDH drug.

Preferably determination of the status of the human is clinically useful. Examples of
5 clinical usefulness include deciding which drug or drugs to administer and/or establishing the
effective amount of the drug or drugs.

Drugs which increase the activity of PDH are of value in a number of disease
conditions, including disease states associated with disorders of glucose utilisation such as
diabetes and obesity, and associated with excessive production of lactate such as encountered
10 in sepsis and other causes of lactic acidemia. Additionally drugs which increase the activity
of PDH may be expected to have utility in diseases where supply of energy-rich substrates to
tissues is limiting such as peripheral vascular disease, coronary failure and certain cardiac
myopathies, muscle ataxia and weakness.

According to another aspect of the present invention there is provided use of a PDH
15 drug in the preparation of a medicament for treating a PDH-mediated disease in a human
diagnosed as having a polymorphism at one or more of positions 857 and 1255 in the PDH E2
gene as defined by the positions in EMBL accession number Y00978.

According to another aspect of the present invention there is provided a
pharmaceutical pack comprising a PDH drug and instructions for administration of the drug to
20 humans diagnostically tested for a polymorphism at one or more of positions 857 and 1255 in
the PDH E2 gene as defined by the positions in EMBL accession number Y00978.

According to another aspect of the present invention there is provided a computer
readable medium comprising at least one novel polynucleotide sequence of the invention
stored on the medium. The computer readable medium may be used, for example, in
25 homology searching, mapping, haplotyping, genotyping or pharmacogenetic analysis or any
other bioinformatic analysis. The reader is referred to Bioinformatics, A practical guide to the
analysis of genes and proteins, Edited by A D Baxevanis & B F F Ouellette, John Wiley &
Sons, 1998. Any computer readable medium may be used, for example, compact disk, tape,
floppy disk, hard drive or computer chips.

30 The polynucleotide sequences of the invention, or parts thereof, particularly those

relating to and identifying the polymorphisms identified herein represent a valuable information source, for example, to characterise individuals in terms of haplotype and other sub-groupings, such as investigation of susceptibility to treatment with particular drugs. These approaches are most easily facilitated by storing the sequence information in a computer readable medium and then using the information in standard bioinformatics programs or to search sequence databases using state of the art searching tools such as "GCC". Thus, the polynucleotide sequences of the invention are particularly useful as components in databases useful for sequence identity and other search analyses. As used herein, storage of the sequence information in a computer readable medium and use in sequence databases in relation to 'polynucleotide or polynucleotide sequence of the invention' covers any detectable chemical or physical characteristic of a polynucleotide of the invention that may be reduced to, converted into or stored in a tangible medium, such as a computer disk, preferably in a computer readable form. For example, chromatographic scan data or peak data, photographic scan or peak data, mass spectrographic data, sequence gel (or other) data.

The invention provides a computer readable medium having stored thereon one or more polynucleotide sequences of the invention. For example, a computer readable medium is provided comprising and having stored thereon a member selected from the group consisting of: a polynucleotide comprising the sequence of a polynucleotide of the invention, a polynucleotide consisting of a polynucleotide of the invention, a polynucleotide which comprises part of a polynucleotide of the invention, which part includes at least one of the polymorphisms of the invention, a set of polynucleotide sequences wherein the set includes at least one polynucleotide sequence of the invention, a data set comprising or consisting of a polynucleotide sequence of the invention or a part thereof comprising at least one of the polymorphisms identified herein.

A computer based method is also provided for performing sequence identification, said method comprising the steps of providing a polynucleotide sequence comprising a polymorphism of the invention in a computer readable medium; and comparing said polymorphism containing polynucleotide sequence to at least one other polynucleotide or polypeptide sequence to identify identity (homology), i.e. screen for the presence of a

polymorphism.

According to another aspect of the present invention there is provided an allelic variant of the human PDH E2 polypeptide having a valine at position 217 and/or an asparagine at position 350 or a fragment thereof comprising at least 10 amino acids provided that the
5 fragment comprises the allelic variant at position 217 and/or position 350.

Fragments of PDH E2 polypeptide are at least 10 amino acids, more preferably at least 15 amino acids, more preferably at least 20 amino acids. The polypeptide of the invention does not encompass naturally occurring polypeptide as it occurs in nature, for example, the polypeptide is at least partially purified from at least one component with which it occurs
10 naturally. Preferably the polypeptide is at least 30% pure, more preferably at least 60% pure, more preferably at least 90% pure, more preferably at least 95% pure, and more preferably at least 99% pure.

According to another aspect of the present invention there is provided an antibody specific for an allelic variant of human PDH E2 polypeptide having a valine at position 217
15 and/or an asparagine at position 350 or a fragment thereof comprising at least 10 amino acids provided that the fragment comprises the allelic variants at position 217 and /or position 350.

Antibodies can be prepared using any suitable method. For example, purified polypeptide may be utilised to prepare specific antibodies. The term "antibodies" is meant to include polyclonal antibodies, monoclonal antibodies, and the various types of antibody
20 constructs such as for example F(ab')₂, Fab and single chain Fv. Antibodies are defined to be specifically binding if they bind the T679M variant of integrin α_4 with a K_a of greater than or equal to about 10^7 M^{-1} . Affinity of binding can be determined using conventional techniques, for example those described by Scatchard et al., *Ann. N.Y. Acad. Sci.*, 51:660 (1949).

Polyclonal antibodies can be readily generated from a variety of sources, for example,
25 horses, cows, goats, sheep, dogs, chickens, rabbits, mice or rats, using procedures that are well-known in the art. In general, antigen is administered to the host animal typically through parenteral injection. The immunogenicity of antigen may be enhanced through the use of an adjuvant, for example, Freund's complete or incomplete adjuvant. Following booster immunisations, small samples of serum are collected and tested for reactivity to antigen.
30 Examples of various assays useful for such determination include those described in:

Antibodies: A Laboratory Manual, Harlow and Lane (eds.), Cold Spring Harbor Laboratory Press, 1988; as well as procedures such as countercurrent immuno-electrophoresis (CIEP), radioimmunoassay, radioimmunoprecipitation, enzyme-linked immuno-sorbent assays (ELISA), dot blot assays, and sandwich assays, see U.S. Patent Nos. 4,376,110 and 4,486,530.

5 Monoclonal antibodies may be readily prepared using well-known procedures, see for example, the procedures described in U.S. Patent Nos. RE 32,011, 4,902,614, 4,543,439 and 4,411,993; *Monoclonal Antibodies, Hybridomas: A New Dimension in Biological Analyses*, Plenum Press, Kennett, McKearn, and Bechtol (eds.), (1980).

10 The monoclonal antibodies of the invention can be produced using alternative techniques, such as those described by Alting-Mees et al., "Monoclonal Antibody Expression Libraries: A Rapid Alternative to Hybridomas", *Strategies in Molecular Biology* 3: 1-9 (1990) which is incorporated herein by reference. Similarly, binding partners can be constructed using recombinant DNA techniques to incorporate the variable regions of a gene that encodes a specific binding antibody. Such a technique is described in Larrick et al., *Biotechnology*, 7:
15 394 (1989).

Once isolated and purified, the antibodies may be used to detect the presence of antigen in a sample using established assay protocols.

The invention will now be illustrated but not limited by reference to the following Examples. All temperatures are in degrees Celsius.

20 In the Examples below, unless otherwise stated, the following methodology and materials have been applied.

AMPLITAQ™ or AMPLITAQ GOLD™ available from Perkin-Elmer Cetus, are used as the source of thermostable DNA polymerase.

25 General molecular biology procedures can be followed from any of the methods described in "Molecular Cloning - A Laboratory Manual" Second Edition, Sambrook, Fritsch and Maniatis (Cold Spring Harbor Laboratory, 1989).

Electropherograms were obtained in a standard manner: data was collected by ABI377 data collection software and the wave form generated by ABI Prism™ sequencing analysis (2.1.2).

Example 1**Identification of Polymorphisms****1. Methods**c-DNA Preparation

- 5 RNA was prepared from lymphoblastoid cell lines from Caucasian donors using standard laboratory protocols (Chomczynski and Sacchi, Anal. Biochem. 162, 156-159, 1987) and used to generate first strand cDNA (Gubler and Hoffman, Gene 25, 263-269, 1983).

Template Preparation

- 10 Templates were prepared by PCR using the oligonucleotide primers and annealing temperatures set out below. The extension temperature was 72° and denaturation temperature 94°; each step was 1 minute. Generally 100 pg cDNA was used in each reaction and subjected to 40 cycles of PCR.

Fragment	Forward Oligo	Reverse Oligo	Annealing Temp	DMSO	MgCl ₂
176-579	176-198	559-579	55°	5%	1.5mM
455-998	455-475	975-998	60°	0%	1.5mM
815-1206	815-836	1185-1206	62°	0%	1mM
1109-1592	1109-1128	1570-1592	62°	0%	1mM
1429-1893	1429-1450	1871-1893	62°	0%	1mM

All positions refer to the positions in EMBL accession number Y00978.

15

For dye-primer sequencing the forward primers were modified to include M13 forward sequence (ABI protocol P/N 402114, Applied Biosystems) at the 5' end of the oligonucleotides.

Dye Primer Sequencing

- 20 Dye-primer sequencing using M13 forward primer was as described in the ABI protocol P/N 402114 for the ABI Prism™ dye primer cycle sequencing core kit with "AmpliTaq FS"™ DNA polymerase, modified in that the annealing temperature was 45° and DMSO was added to the cycle sequencing mix to a final concentration of 5%.

The extension reactions for each base were pooled, ethanol/sodium acetate precipitated, washed and resuspended in formamide loading buffer.

4.25% Acrylamide gels were run on an automated sequencer (ABI 377, Applied Biosystems).

5

2. Results

Novel Polymorphisms

<i>Position</i>	<i>Published allele</i>	<i>Variant allele</i>	<i>amino acid change</i>	<i>RFLP</i>	<i>Allele Frequency</i>
857	T	C	Ala-Val	+ Hha I	11/40
1255	G	A	Asp-Asn	- Cla I	17/38

10

All positions refer to the positions in EMBL accession number Y00978.

Frequency is the allele frequency of the variant allele in control subjects.

Example 2

15 *Analysis of the sequences of PDH E2 disclosed in public databases*

Two cDNA sequences encoding PDH E2 have been submitted to public databases under accession numbers Y00978 and J03866. Y00978 identifies an open reading frame (ORF) which extends previously published sequence at the 5' end but does not identify an
 20 ATG start site. J03866 contains more 5' sequence than Y00978 and identifies an ATG start site. However, the 5' sequence of J03866 does not match that of Y00978.

We used sequence regions common to Y00978 and J03866 to search public and proprietary sequence databases and were able to obtain extended sequence at the 5' end. The extended sequence identified the start of an open reading frame which was in frame with the
 25 ORFs reported within both Y00978 and J03866. The initiating ATG corresponded to position 211 of Y00978, position 843 of J03866.

Sequence at positions 1-698 of J03866 did not match the extended sequence determined in our analysis. Sequence J03866 1-698 was found to be identical to murine sulphatase A. Furthermore, we were unable to amplify the predicted PCR product sizes from human lymphoblastoid cell line cDNA using PCR primers designed from sequence J03866 1-
5 698, providing further confirmation that sequence J03866 1-698 is incorrect. J03866 also differs from Y00978 by three missing nucleotides.

Sequence analysis of lymphoblastoid cell line cDNA agreed with the sequence of Y00978. In conclusion, we believe that sequence Y00978 is correct and that the ATG start site corresponds to position 211 of Y00978.

Application No: NEW

Pillsbury WINTHROP

Inventor: SMITH

Date: 1/18/01

Client & Ref. #: ASTRAZENECA

CL.# 9901 M# 276522